

# EXTENSION

Institute of Food and Agricultural Sciences

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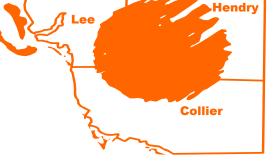
# Flatwoods Citrus

Vol. 26, No. 8

August 2023

Dr. Mongi Zekri Multi-County Citrus Agent, SW Florida





Mongi Zekri

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### **September 2023 Zoom Citrus Workshops**

Date and time: Thursday, September 14, 2023, 10:00 AM - 11:00 AM

**Title:** "Breeding citrus for tolerance to HLB"

Speaker: Dr. Jude Grosser, Professor in cell plant genetics at the UF-IFAS

Citrus Research & Education Center in Lake Alfred Zoom link will be provided by the end of this month

Date and time: Thursday, September 28, 2023, 10:00 AM – 11:00 AM

**<u>Title:</u>** "Silicon - A beneficial nutrient for citrus production"

Speaker: **Dr. Muhammad Adnan Shahid**, Assistant Professor of Horticulture at the UF-IFAS North Florida Research and Education Center in Quincy Discussions will include:

- -What is the silicon and its benefits
- -Different sources of silicon fertilizer
- -Application methods of silicon and application rate
- -Use of silicon in citrus production

Zoom link will be provided next month

Coordinator: Dr. Mongi Zekri, UF-IFAS, maz@ufl.edu

1 CEU for pesticide license renewal

1 CEU for certified crop advisors

### **CEUs for pesticide license renewal**

Earn CORE CEUs online through articles written by UF-IFAS Citrus Extension Agents in the Citrus Industry magazine <a href="http://citrusindustry.net/ceu/">http://citrusindustry.net/ceu/</a>

The following series of articles and quizzes are available with their expiration dates noted:

- 2023 #3: A Guide to Safe, Effective Pesticide Use (7/31/24)
- 2023 #2 What To Do When You've Been Exposed to a Pesticide (4/30/24)
- 2023 #1: Key Terms to Know When Using Pesticides (1/31/24)
- 2022 #4: Making Sense of Pesticide Formulations (10/31/23)

Each article grants one General Standards (Core) CEU when submitted and approved toward the renewal of a Florida Department of Agriculture and Consumer Services restricted-use pesticide license.

### Florida Citrus Production Guide

https://crec.ifas.ufl.edu/resources/production-guide/

The objective of the Florida Citrus Production Guide is to assist citrus growers in the identification of pest management options and the selection of appropriate control measures. This publication should serve as a reference once it has been determined that control measures might be warranted. It is not intended to replace pesticidal product labels which contain important usage information and should be immediately accessible for reference. Violations of directions for use printed on the label are against State and Federal laws. Care should be taken to select only those treatments best suited for control of the specific pest(s) identified as requiring suppression. Products listed in all tables have been shown to be efficacious, non-phytotoxic to citrus, and relatively safe on non-target arthropods and microorganisms when used as directed. However, it is important to realize that results may not be consistent under different environmental, application, and tank mix conditions.

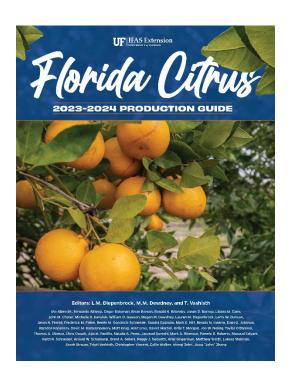
### **PRODUCTION GUIDE MENU**

- General
- Horticultural Practices
- Mites, Insects & Nematodes
- Diseases
- Weeds
- Pesticides

If you did not pick up your hard copy of the newly updated Florida Citrus Production Guide at the Citrus & Specialty Crop Expo, you can find the electronic version online

https://crec.ifas.ufl.edu/resources/pr oduction-guide/

If you need hard copies, you can get them free from your Citrus Extension Agent or from the Citrus Research & Education Center in Lake Alfred and the Southwest Florida Research and Education Center in Immokalee.



# 2023-2024 Citrus Production Guide Available at Florida Citrus Expo

An updated perennial resource for Florida commercial citrus growers is available to support their efforts to produce a successful citrus harvest

# PUBLISHED ON JULY 24, 2023

The 2023-2024 publication includes the latest technical advice on citrus greening disease management, tree nutrition, grove management and other aspects of Florida citrus cultivation. (Courtesy Photo)

LAKE ALFRED, Fla. — An updated perennial resource for Florida commercial citrus growers is available to support their efforts to produce a successful citrus harvest. The 2023-2024 UF Institute of Food and Agricultural Sciences Citrus Production Guide will be available to growers at the August Florida Citrus Expo in Tampa and through their local UF/IFAS Extension citrus agents afterwards.

The document is a comprehensive reference meant to assist growers in development and management of citrus groves in Florida, said Tripti Vashisth, a UF/IFAS horticulture associate professor and a member of the guide's editorial team.

Additions this year include new fertilizer recommendations for some nutrients that have been recently changed and information on the plant growth regulator gibberellic acid (GA). Information on pesticides registered for use in Florida citrus is also updated in every new edition.

Topics covered in the guide include planting, irrigation, fertilization, weed control, insect management and disease management, she said. For a grove management practice to be recommend in the guide, a minimum of two years of field validated data is required. About 50 UF/IFAS personnel contributed to the latest edition.

"Each year, we update the guide with new information as it becomes available. This is a fundamental component of the UF/IFAS faculty mission to continue to meet the needs of our growers as we learn about and/or develop new tools and strategies for citrus production," Vashisth said.

Hard copies of the updated guide will be available at the UF/IFAS booth at the Florida Citrus Expo held at the Florida State Fairgrounds in Tampa, August 16 – 17. After that, they will be at local UF/IFAS Extension county offices (please see chart) in citrus growing counties.

For a directory of citrus agents, visit <a href="http://citrusagents.ifas.ufl.edu/locate/index.shtml">http://citrusagents.ifas.ufl.edu/locate/index.shtml</a>. Another way to access the guide is on-line through the UF/IFAS Extension Electronic Data Information Source library at the EDIS website. It will also be available at the UF/IFAS Citrus Research and Education Center and the <a href="mailto:cirtrusresearch.ifas.ufl.edu">cirtrusresearch.ifas.ufl.edu</a> websites.



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If you would like to be among them, please contact me at 863 674 4092 or maz@ufl.edu

### 2023 Citrus & Specialty Crop Expo Seminar Program

SEMINAR SCHEDULE TIMES ARE SUBJECT TO CHANGE.

Wednesday, August 16, 2023

7:30 a.m. - 3 p.m. - Registration at Trade Show Entrance

7:30 a.m. - 9:30 a.m. - Breakfast in Trade Show Hall

8:00 a.m. - 5:30 p.m. - Trade Show Open

Visit exhibitor booths!

3:30 p.m. - 5:30 p.m. - Networking Reception

Hit the tradeshow floor and grab a cocktail!

### MORNING SESSION (9:00 A.M. - 12:05 P.M.)

**General Session** 

9:00 - Welcome, announcements and introduction

9:15 - Citrus Research and Development Foundation (CRDF) Update

Rick Dantzler, CRDF

9:30 - New Markets for Ecosystems Services

Moderator: Ernie Shea, Solutions from the Land; David Hill, Southern Hill Farms and Joel Harley, University of Florida Department of Electrical and Computer Engineering

10:15 - Statewide Best Management Practices Priorities

West Gregory, Florida Department of Agriculture and Consumer Services (FDACS),

Office of Agricultural Water Policy

10:35 - Agritourism Opportunities in Florida

Michelle Welch, Florida Agritourism Association

10:55 - Crop Insurance Enhancements for Citrus and Specialty Crop Growers

Bryan McQuary, U.S. Department of Agriculture, Risk Management Agency

11:15 - Updates to Farm Service Agency (FSA) Disaster Programs

Kathy Sayers, USDA FSA

11:35 - Keynote Address

Florida Commissioner of Agriculture Wilton Simpson, FDACS

12:05 - Lunch

AFTERNOON SESSION (1:50 P.M. - 4:35 P.M.)

Citrus Seminars

CEUS AND CCAS AVAILABLE

1:50 - Welcome/Sponsor Introduction

2:00 - An Update From UF/IFAS

Rob Gilbert, UF/IFAS

2:15 - Progress Developing New Citrus Varieties Using CRISPR and Other Biotech Approaches

Michael Rogers, UF/IFAS

2:45 - IPCs Work. What Comes Next?

Fernando Alferez, UF/IFAS

2:55 - Optimizing Irrigation and Fertilization in HLB-Affected Trees

Davie Kadyampakeni, UF/IFAS

3:05 - Employing Plant Growth Regulators to Improve Canopy Health

Tripti Vashisth, UF/IFAS

3:15 - The Latest on Trunk Injection HLB Therapy

Ute Albrecht, UF/IFAS

3:25 - Psyllid Research and Management Update

Lukasz Stelinski, UF/IFAS

3:35 - Discussion Panel and Q&A With Speakers

4:35 - Visit Trade Show and Networking Reception

Vegetable and Specialty Crop Seminars

CEUS AND CCAS AVAILABLE

2:00 - Welcome/Sponsor Introduction

2:10 - Planning for the Next Vegetable Season: Thrips parvispinus Management Anna Mészáros, UF/IFAS

2:20 - Managing Pepper Weevil: Considerations, Suppression and Screen Barrier Protection

Jawwad Qureshi, UF/IFAS

2:40 - Increasing Efficiency and Optimizing Economic Return in Relay and Double Cropping Systems

Nathan Boyd, UF/IFAS

3:00 - Optimizing Summer Fallow Practices to Enhance Weed Suppression in Vegetables

Ramdas Kanissery, UF/IFAS

3:20 - Team Presentation: Nutrient BMP Research for Beans, Potatoes and Tomatoes Thomas Obreza, Sanjay Shukla, Shinsuke Agehara, Christian Christensen, Kimberly Morgan, UF/IFAS

4:00 - Visit Trade Show and Networking Reception

Thursday, August 17, 2023

7:30 a.m. - 3 p.m. - Registration at Trade Show Entrance

7:30 a.m. - 9:30 a.m. - Breakfast in Trade Show Hall

8:00 a.m. - 2:00 p.m. - Trade Show Open

Visit exhibitor booths!

MORNING SESSION (8:50 A.M. - 12:30 P.M.)

Citrus Seminars

CEUS AND CCAS AVAILABLE

8:50 - Welcome/Sponsor Introduction

9:00 - Artificial Intelligence for Precision Grove Management

Yiannis Ampatzidis, UF/IFAS

9:20 - Transgene-Free CRISPR Citrus Genome Editing

Nian Wang, UF/IFAS

9:40 - Understanding and Managing Soil Health of Citrus

Sarah Strauss, UF/IFAS

10:00 - Enhancing Weed Control: Latest Updates for Mature and Young Groves Ramdas Kanissery, UF/IFAS

10:20 - Multi-Year Comparison of New Tools to Support Establishment of Young Groves

Lauren Diepenbrock, UF/IFAS

10:40 - Supporting Establishment of Young Groves and Keeping Diseases at Bay Megan Dewdney, UF/IFAS

11:00 - US Early Pride and New Selections for Use in CUPS

Matt Mattia, USDA Agricultural Research Service

11:20 - Sweet Oranges from the UF/IFAS Citrus Breeding Team

John Chater, UF/IFAS

11:40 - Strategies to Mitigate HLB in Groves

**Grower Panel** 

12:30 - Visit Trade Show/Lunch

Vegetable and Specialty Crop Seminars

CEUS AND CCAS AVAILABLE

9:00 - Sponsor Introduction

9:15 - Welcome Comments

Andra Johnson, UF/IFAS

9:20 - Optimizing Irrigation Management to Maximize Crop Health

Craig Frey, UF/IFAS

9:40 - Florida Stakeholder Engagement Program: Optimizing Agricultural Water,

Nutrient and Economic Efficiency

Vivek Sharma, UF/IFAS

10:00 - Sustainable Vegetable Production in Florida

Pavlos Tsouvaltzis, UF/IFAS

10:20 - Characterization of Management of Cabbage Black Rot in Florida

Renzo Ramirez, UF/IFAS

10:40 - Monitoring Insecticide Susceptibility of Whitefly (Bemisia tabaci MEAM1) in Florida

Hugh Smith and Marcelo Dimase, UF/IFAS

11:00 - FAWN Data and Tools for Decision Making: New Enhancements

Rick Lusher, UF/IFAS

11:20 - Keeping On Top of Nematode Pressure

Johan Desaeger, UF/IFAS

11:40 - Lunch

### **PHYTOPHTHORA**

Foot rot results from infection of the scion near the ground level, producing bark lesions, which extend down to the budunion on resistant rootstocks.



Crown rot results from infection of the bark below the soil line when susceptible rootstocks are used. Root rot occurs when the cortex of fibrous roots is infected, turns soft and appears water-soaked.

Fibrous roots slough their cortex leaving only white thread-like stele.



When managing Phytophthora-induced diseases, consider integration of cultural practices (e.g., disease exclusion through use of Phytophthora-free planting stock, resistant rootstocks, proper irrigation practices) and chemical control methods.

<u>Cultural practices</u>. Field locations not previously planted with citrus are free of citrus-specific *P. nicotianae*. Planting stock should be tested free of Phytophthora in the nursery and inspected for fibrous root rot in the nursery or grove before planting. In groves with a previous history of foot rot, consider use of Swingle citrumelo for replanting. Swingle citrumelo is resistant to foot rot and roots do not support damaging populations once trees are established.

Cleopatra mandarin should be avoided because it is prone to develop foot rot when roots are infected in the nursery or when trees are planted in flatwoods situations with high or fluctuating water tables and fine-textured soils. Trees should be planted with the budunion well-above the soil line and provided with adequate soil drainage. Overwatering, especially of young trees, promotes buildup of populations in the soil and increases risk of foot rot infection. Prolonged wetting of the trunk, especially if tree wraps are used on young trees, should be avoided by using early to midday irrigation schedules. Control of fire ants prevents their nesting under wraps and causing damage to tender bark.

Sampling for *P. nicotianae*. Population densities of the fungus in grove soils should be determined to assist in decisions to treat with fungicides. Soil samples containing fibrous roots should be collected during the spring through fall (March to November) from undercanopy within the tree dripline. Individual small amounts of soil from 20 to 40 locations within a 10-acre area are composited into one resealable plastic bag to retain soil moisture. Samples must be kept cool but not refrigerated for transport to the analytical laboratory. Currently, populations in excess of 10 to 15 propagules per cm³ soil are considered damaging. The same soil sample could be

tested for populations of nematodes, to assess whether they occur at damaging levels.

<u>Chemical control.</u>

Use of fungicides in young groves should be based on rootstock susceptibility, likelihood of Phytophthora infestation in the nursery, and history of Phytophthora disease problems in the grove. For susceptible rootstocks, such as Cleopatra mandarin and sweet orange, fungicides may be applied to young trees on a preventive basis for foot rot. For other rootstocks, fungicide treatments should commence when foot rot lesions develop. The fungicide program for foot rot should be continued for at least one year for tolerant rootstocks, but may continue beyond for susceptible stocks.



In mature groves, the decision to apply fungicides for root rot control is based on yearly soil sampling to indicate whether damaging populations of *P. nicotianae* occur in successive growing seasons. Time applications to coincide with periods of susceptible root flushes in late spring and late summer or early fall.

Soil application methods with fungicides should be targeted to under canopy areas of highest fibrous root density. To avoid leaching from the root zone, soil-applied fungicides should not be followed by excessive irrigation.



Recommended Chemical Controls for Phytophthora Foot Rot and Root Rot include Aliette, Phostrol, ProPhyt, Ridomil, UltraFlourish, and Copper.

For more details, go to: <a href="https://edis.ifas.ufl.edu/pdf/CG/CG009/CG009-Dtzflnc7al.pdf">https://edis.ifas.ufl.edu/pdf/CG/CG009/CG009-Dtzflnc7al.pdf</a>

### **GREASY SPOT**

Management of greasy spot must be considered in groves intended for processing and fresh market fruit. Greasy spot is usually more severe on leaves of grapefruit, pineapples, Hamlins, and tangelos than on Valencias, Temples, Murcotts, and most tangerines and their hybrids.

Greasy spot spores germinate on the underside of the leaves and the fungus penetrates through the stomates (natural openings on lower leaf surface). Warm humid nights and high rainfall, typical of Florida summers, favor infection and disease development.





On processing Valencias, a single spray of oil (5-10 gal/acre) or copper + oil (5 gal/acre) should provide acceptable control when applied from mid-May to June. With average quality copper products, 2 lb of metallic copper per acre usually provide adequate control. The strobilurin fungicides (Abound, Gem, Headline or Quadris), as well as Enable 2F, are also suitable with or without petroleum oil. On early and midseason oranges and grapefruit for processing, two sprays may be needed especially in the southern part of the state where summer flushes constitute a large portion of the foliage. Two applications also may be needed where severe defoliation from greasy spot occurred in the previous year. In those cases, the first spray should be applied from mid-May to June and the second soon after the major summer flush has expanded. Copper fungicides provide a high degree of control more consistently than oil sprays. Control of greasy spot on late summer flushes is less important than on the spring and early summer growth flushes since the disease develops slowly and defoliation will not occur until after the next year's spring flush. Thorough coverage of the underside of leaves is necessary for maximum control of greasy spot, and higher spray volumes and slower tractor speeds may be needed than for control of other pests and diseases.



The program is essentially the same for fresh fruit. That is, a fungicide application in May-June and a second in July should provide control of rind blotch.

A third application in August may be needed if rind blotch has been severe in the grove. Petroleum oil alone is less effective than other fungicides for control of greasy spot rind blotch (GSRB). Heavier oils (455 or 470) are more effective for rind blotch control than are lighter oils (435).

Copper fungicides are effective for control of GSRB, but may result in fruit spotting especially if applied at high rates in hot, dry weather or if applied with petroleum oil. If copper fungicides are applied in summer, they should be applied when temperatures are moderate, at rates no more than 2 lb of metallic copper per acre, without petroleum oil or other additives, and using spray volumes of at least 125 gal/acre. Enable 2F can be applied for greasy spot control at any time but is especially indicated in mid to late summer for rind blotch control.

The strobilurin fungicides (Abound, Gem, Headline, Pristine or Quadris Top) or Enable 2F can be applied at any time to all citrus and provide effective control of the disease on leaves or fruit. Use of a

strobilurin (Abound, Gem, Headline, Pristine or Quadris Top) is especially indicated in late May and early June since it will control both melanose and greasy spot and avoids potential fruit damage from the copper fungicides at that time of year. A strobilurin fungicide should not be applied more than once a year for greasy spot control. Addition of petroleum oil increases the efficacy of these products.

### Processed fruit

### **July-August**

- Petroleum oil (455, 470) 5-10 gal
- Cu fungicides 2-4 lb metal
- Abound, Gem, Headline + 5 gal oil
- Pristine
- Amistar Top
- Enable

### •Fresh fruit

### **July-August**

- Petroleum oil (455, 470) 10 gal
- Cu fungicides < 2 lb metal
- Abound, Gem, Headline + 5 gal oil
- Pristine
- Amistar Top
- Enable 8 oz

### For more information on greasy spot, go to:

https://edis.ifas.ufl.edu/pdf/CG/CG018/CG018-Dmqxycvuom.pdf

# LEAF AND SOIL SAMPLING AND ANALYSES

Optimum growth and yield of high quality fruit cannot be obtained without adequate nutrition. The most successful fertilizer program should be based on tissue analysis, knowledge of soil nutrient status through soil analysis combined with university recommendations. The deficiency or excess of an element will cause disturbance in plant metabolism and lead to poor performance.



### Plant analysis

Used in conjunction with other data and observations, tissue analysis aids in evaluating the nutrient elements of the soil-plant system. It has proven useful in confirming nutritional deficiencies, toxicities or imbalances, identifying "hidden" toxicities and deficiencies where visible symptoms are not manifested, and evaluating the effectiveness of fertilizer programs.

### **Leaf Sampling**

For reliable results and useful interpretation of lab analysis reports, citrus growers, production managers, and consultants must follow the proper procedures for leaf sampling and sample handling because improperly collected leaf samples will provide misleading information about the nutritional status of the trees and the fertilizer programs.

Considerable care is needed in taking samples. Chemical analysis values can only be useful if the samples obtained are representative of the blocks they were taken from. The proper sampling, preparation and handling would affect the reliability of the chemical analysis, data interpretation, nutritional recommendations, and adjustment of fertilizer programs.

Leaf samples must also be taken at the proper time because nutrient levels within leaves are continually changing. However, leaf mineral concentrations of most nutrients are relatively stable within 4 to 6 months after emergence of the spring flush. Therefore, for mature tree blocks, the best time would be in <u>July and August</u> to collect four- to six-month-old spring flush leaves. If taken later in the season, the summer flush would probably be confused with the spring flush.

Each leaf sample should consist of about 100 leaves taken from non-fruiting twigs of 15- 20 uniform trees of the same variety and rootstock, and under the same fertilizer program. Clean brown paper bag should be used. Information sheets from the testing lab should be completed for each sample as this information helps when interpreting the results. The sample bag and the corresponding information sheet should each be carefully labeled with the same identity so that samples and sheets can be matched in the laboratory.

### Sampling techniques for leaves

- ♦ Immature leaves should be avoided because of their rapidly changing composition.
- ♦ Abnormal-appearing trees, trees at the edge of the block and trees at the end of rows should not be sampled because they may be coated with soil particles and dust or have other problems.
- ♦ Do not include diseased, insect damaged, or dead leaves in a sample. Use good judgment.

♦ Select only one leaf from a shoot and remove it with its petiole (leaf stem).

### **Diagnosing growth disorders**

- ♦ Collect samples from both affected trees as well as normal trees.
- ♦ Trees selected for sampling should be at similar stage of development and age.
- ♦ Whenever possible, confine the sampling area to trees in close proximity to each other.

### **Handling of leaf samples**

- ◆ Samples should be collected in clean paper bags and clearly identified.
- ♦ They should be protected from heat and kept dry and cool (stored in portable ice chests), and placed in a refrigerator for overnight storage if they cannot be washed and oven dried the same day of collection.
- ♦ For macronutrient analysis, leaves usually do not need to be washed.
- ♦ Leaves should be dried in a ventilated oven at 60-70°C.

### **Preparation for analysis**

- ♦ Leaves that have been recently sprayed with micronutrients for fungicidal (Cu) or nutritional (Mn, Zn) purposes should not be analyzed for those micronutrients because it is unlikely to remove all surface contamination from sprayed leaves.
- ♦ For accurate Fe and B or other micronutrient determination, samples would require hand washing, which is best done when leaves are still in a fresh condition.

### Soil analysis

Soil analysis is an important method for gaining basic information regarding the chemical status of the soil. Soil analysis is particularly useful when conducted over several years so that trends can be seen.

Unlike leaf analysis, there are various methods and analytical procedures of soil analysis used by laboratories. In Florida, soil tests for the relatively mobile and readily leached elements such as N and K are of no value. Soil tests are mainly important for pH, P, Mg, Ca, and Cu. For

Florida sandy soils, using the Mehlich-1 or double acid (hydrochloric acid + sulfuric acid) extraction procedure adopted by the University of Florida analytical lab, 40-60 lbs/acre (20-30 ppm) of P, 70-120 lbs/acre (35-60 ppm) of Mg, 500-800 lbs/acre (250-400 ppm) of Ca, and 5-10 lbs/acre (2.5-5 ppm) of Cu are considered adequate for citrus. A Ca:Mg ratio of 7:1 seems desirable and ratios of higher than 10 may induce Mg deficiency problems. Copper levels higher than 50 lbs/acre may be toxic to citrus trees if the soil pH is below 6.

### Soil sampling

The accuracy of a fertilizer recommendation depends or how well the soil sample on which the recommendation was based represents the area of the grove. In Florida, if soil samples were to be collected once a year, the best time would be at the end of the summer rainy season and prior to fall fertilization, usually during September and October. However, soil sampling may be conducted at the same time as leaf sampling to save time and reduce cost.

Standard procedures for proper sampling, preparation and analysis have to be followed for meaningful interpretations of the test results and accurate recommendations. Each soil sample should consist of 15-20 soil cores taken at the dripline of 15-20 trees within the area wetted by the irrigation system to a depth of 6 inches. The area sampled should be uniform in terms of soil and tree characteristics and correspond to the area from which the leaf sample was taken. Individual cores should be mixed thoroughly in a plastic bucket to form a composite sample. Subsample of appropriate size should be taken from the composite mixture and put into labeled paper bags supplied by the lab. Soil samples should be air-dried but not oven-dried before shipping to the testing laboratory for analysis.

### Conclusion

Tissue and soil analyses are a powerful tool for confirming nutrient deficiencies, toxicities and imbalances, identifying "hidden hunger," evaluating fertilizer programs, studying nutrient interactions. However, if initial plant and soil sampling, handling, and analysis of the sample were faulty, the results would be misleading.

If properly done, tissue and soil analyses can point the way toward more economical and efficient use of fertilizer materials, avoiding excessive or inadequate application rates.

For more details, consult UF-IFAS publication SL 253, "Nutrition of Florida Citrus Trees," at http://edis.ifas.ufl.edu/pdffiles/SS/SS47800.pdf

# Standard Table for Assessing Nutritional Status and Adjusting Fertilizer Programs for Citrus

Leaf analysis standard for assessing current nutrient status of citrus trees based on concentration of mineral elements in 4- to 6-month-old-spring-cycle leaves from non-fruiting terminals.

Element	Deficient less than	Low	Satisfactory	High	Excess more than
Nitrogen (N) (%)	2.2	2.2-2.4	2.5-2.8	2.9-3.2	3.3
Phosphorus (P) (%)	0.09	0.09-0.11	0.12-0.17	0.18-0.29	0.30
Potassium (K) (%)	0.7	0.7-1.1	1.2-1.7	1.8-2.3	2.4
Calcium (Ca) (%)	1.5	1.5-2.9	3.0-5.0	5.1-6.9	7.0
Magnesium (Mg) (%)	0.20	0.20-0.29	0.30-0.50	0.51-0.70	0.80
Sulfur (S) (%)	0.14	0.14-0.19	0.20-0.40	0.41-0.60	0.60
Chlorine (Cl) (%)			less than 0.5	0.5-0.7	0.7
Sodium (Na) (%)			less than 0.2	0.2-0.5	0.5
Iron (Fe) (ppm)	35	35-59	60-120	121-200	250
Boron (B) (ppm)	20	20-35	36-100	101-200	250
Manganese (Mn) (ppm)	18	18-24	25-100	101-300	500
Zinc (Zn) (ppm)	18	18-24	25-100	101-300	300
Copper (Cu) (ppm)	4	4-5	6-16	17-20	20
Molybdenum (Mo) (ppm)	0.06	0.06-0.09	0.1-1.0	2-50	50

# IRRIGATION, NUTRITION AND FRUIT QUALITY

Florida has the highest citrus fruit quality standards in the world. Fruit quality factors include juice content, soluble solids and acid concentrations, soluble solids-acid ratio, fruit size, and color. Florida citrus growers know that quality factors differ for the fresh and processing markets. For example, fruit size, shape, color, and maturity date are most important for fresh fruit, but high juice content and soluble solids are desired for processing fruit. Fruit quality is affected by several factors including cultivar, rootstock, climate, soil, pests, irrigation, and nutrition.

The effects of irrigation and nutrition on fruit quality are very important and should be understood and taken into consideration by citrus growers and production managers to increase their profitability and enhance their sustainability and competitiveness on a worldwide basis. In general, excessive irrigation and nutrition reduce fruit quality. Therefore, balanced nutrition with sound irrigation scheduling based on **IFAS** recommendations should be a high priority management practice for every grower. Citrus trees require a properly designed, operated, and maintained water management system and a balanced nutrition program formulated to provide specific needs for maintenance and for expected yield and fruit quality performance. Irrigation contributes to the efficiency of fertilizer programs. Adequately watered and nourished trees grow stronger, have better tolerance to pests and stresses, yield more consistently, and produce good quality fruit. On the other hand, excessive or deficient levels of watering or fertilization will result in poor

fruit quality. The most important management practices influencing fruit quality are irrigation and nitrogen, phosphorus, potassium, and magnesium nutrition. However, when any nutrient element is severely deficient, fruit yield and fruit quality will be negatively altered. Trends in fruit quality response to high nutrition and irrigation are described and summarized below.

### Nitrogen (N)

- □ Increases juice content and color, total soluble solids (TSS), and acid content.
- Increases soluble solids per box and per acre. However, excessive N, particularly with inadequate irrigation, can result in lower yields with lower TSS per acre.
- Decreases fruit size and weight.
- □ Increases peel thickness and green fruit at harvest.
- □ Increases incidence of creasing and scab but decreases incidence of peel blemishes such as wind scar, mite russeting, and rind plugging.
- □ Reduces stem-end rot incidence and green mold of fruit in storage.

### Phosphorus (P)

- Reduces acid content, which increases soluble solids-acid ratio.
   Phosphorus rates have no effect on soluble solids per box but may increase soluble solids per acre due to increase in fruit production in soils that are low in P.
- □ Increases number of green fruit but reduces peel thickness.
- □ Increases expression of wind scar but reduces that of russeted fruit.

### Potassium (K)

 Potassium produces mostly negative effects on juice quality except soluble solids per acre.
 Potassium increases fruit

- production therefore producing more soluble solids per acre.
- □ Decreases juice content, soluble solids, ratio, and juice color.
- □ Increases acid content.
- □ Increases fruit size, weight, green fruit and peel thickness.
- □ Reduces incidence of creasing and fruit plugging. In storage, reduces stem-end rot.

### Magnesium (Mg)

- Slightly increases soluble solids, soluble solids-acid ratio, soluble solids per box and soluble solids per acre.
- Slightly increases fruit size and weight but decreases rind thickness.

### **Irrigation**

□ Increases juice content and soluble solids-acid ratio.

- Reduces soluble solids and acid contents. Soluble solids per box will decrease, but soluble solids per acre may increase due to yield increase.
- □ Increases fruit size and weight, increases green fruit at harvest, but decreases rind thickness.
- ☐ Increases incidence of blemish from wind scar, scab and *Alternaria* brown spot, but reduces rind plugging.
- Reduces stem-end rot incidence but increases incidence of green mold in storage.

Specific effects on juice and external fruit qualities are summarized in the Table below. This summary is based on numerous field experiments conducted over many years. Most of these effects were consistently observed, but some of them appear to depend on local conditions and growing regions.

# EFFECTS OF MINERAL NUTRITION AND IRRIGATION ON FRUIT QUALITY

Variable	N	P	K	Mg	Irrigation
Juice Quality					
juice content	+	0	-	0	+
soluble solids (SS)	+	0	-	+	-
acid (A)	+	-	+	0	-
SS/A ratio	-	+	-	+	+
juice color	+	0	-	?	0
solids/box	+	0	-	+	-
solids/acre	+	+	+	+	+
External Fruit Quality					
Size	-	0	+	+	+
Weight	-	0	+	+	+
green fruit	+	+	+	0	+
peel thickness	+	-	+	-	-

Increase (+), Decrease (-), No change (0), No information (?).

### **A**LGAE



# Algae are in the plant kingdom, but maybe they're not really plants!

In Florida's freshwaters, algae are what make the water green, or even "slimy". However, green water is not necessarily undesirable, and neither are algae. In fact, algae are essential to the ecosystem and to life as we know it, and must be treated with respect.

Algae are a diverse group of organisms, which survive in all different types of habitats. They range in size from microscopic to meters in length and in complexity from single-celled to complex organisms that would rival even large plants. Though these organisms may look like the true, "higher", plants, they are anything but, since they do not have roots or true stems and leaves.

Algae are one of the first steps of the food web. There are microscopic algae, like phytoplankton, and there are macroalgae, algae that can be seen by the naked eye. Algae occur naturally in all types of systems and may be considered indicators of ecosystem condition. Even the mere presence of a species can give an indication of the amount and type of nutrients that run through the system. Algae provide food for all types of animals, including fish, insects, mollusks, zooplankton (microscopic animals), and humans.

### What causes an algae bloom?

At times algae can grow so quickly and densely that they form a "bloom". Many people don't like the "look" of a bloom, though blooms can be a natural occurrence. Blooms are not necessarily green, though that is the most common color. They can be blue-green, brown, red, and even violet.



Some blooms turn the water a certain color; this is usually a bloom associated with phytoplankton (microscopic algae). Other blooms form clumps or mats that float on top of the water, or that grow attached to the bottom or to plants. Still others can form dense mats that cover the water surface. Algae need nutrients, such as nitrogen and phosphorous, and light to grow. The level of growth or productivity is often dependent on the amount of nutrients in a system. There is a classification for productivity of a system; it ranges from oligotrophic (low productivity and nutrients) to hypereutrophic (very high nutrients). Also, since algae need light to photosynthesize, how far light penetrates the water is also another limiting factor.

Blooms can have far reaching effects on the environment. Some can become so dense they can ultimately cause a problem with low oxygen levels. A decrease in oxygen causes hypoxia (low oxygen) or anoxia (no oxygen) and the other organisms in the water that need oxygen to survive, such as fish, become stressed and may die. Other

blooms may release toxins that can be harmful to animals.

There is a general consensus that rapidly growing human development, and increased human use and disposal of nutrients over the past few centuries, has increased the frequency and intensity of algal blooms in many regions of the world. This has created a global effort to control harmful blooms.

### **Controlling blooms**

### The most direct way to control

blooms is to reduce the availability of nutrients. Most water management organizations throughout the world are actively pursuing a variety of nutrient control strategies. However, for some aquatic ecosystems nutrient control is impractical, ineffective or simply too costly. For some cases, chemical or biological treatments can be helpful alternatives.

### **Chemical Treatments**

Copper sulfate (bluestone) and chelated copper compounds such as Cutrine-Plus, Algae Pro, and K-TEA, as well as Endothall are common chemical treatments used to kill algae. Chemical compounds that shade out the light for algae growth, e.g. Aquashade, are also used to control blooms. Each chemical has its own restrictions and toxicity to animals. Read the directions carefully before application.

### **Biological Treatments**

The main biological treatment that is employed today is the use of various carp fish species to control submersed and floating algae. **Grass carp** (*Ctenopharyngodon idella*) is mainly used for aquatic weeds and attached submersed algae, such as *Nitella* sp., and *Chara* sp. Where they do not prefer filamentous algae to eat, grass carp will eat *Lyngbya*. The **silver carp** (*Hypophthalmichthys molitrix*) has been shown to be an effective treatment

for controlling filamentous algae, including blue-green algae.

Both species are non-native species and there are many restrictions to employing them as a means of weed control; some states prohibit their use altogether. When they are allowed, the use is restricted to **triploid carp**. Triploid carp have an extra set of chromosomes that render the fish sterile, therefore prohibiting a population explosion if the fish escapes into an uncontrolled area.

### **Physical Treatments**

Physical treatments for algae in ponds include aeration and airlifts. While aeration does not kill or remove algae from the water, it oxygenates and stirs the water column, and can create conditions to shift from toxic and smelly blue-green algae to preferred green algae species. The resultant algal population is usually not as dense or as toxic to other organisms in the ponds.

### **Mechanical Treatments**

Harvesters are sometimes used to skim dense mats of blue-green lyngbya alga from the surface of lakes and rivers. Lyngbya normally grows in dense mats at the bottoms of nutrient enriched lakes. These mats produce gasses during photosynthesis that often causes the mats to rise to the surface. At the surface, winds pile the algal mats against shorelines or in navigation channels; these mats can be several acres in size. Managers have developed a process called "grubbing" whereby harvesting machines lift the mats off of submersed plants such as native eelgrass, without cutting the eelgrass. By removing the blanket of lyngbya from the eelgrass, the plants grow and expand. Eelgrass is an important food source for manatees in the Crystal and Homossassa Rivers.

# EFFECT OF WATER pH ON EFFICACY OF PESTICIDES



Successful citrus growers should check the soil pH of their groves yearly and do their best to adjust it for better fertilizer efficiency, tree growth, and fruit production. Soil pH is usually increased by liming and decreased by applying sulfur or acid-forming fertilizers. The pH indicates whether the solution or media is acidic or basic (alkaline). The pH scales goes from 0 to 14, where 7 indicates neutrality. Values less than 7 indicate acidic solutions and values greater than 7 indicate a basic condition. Most of Florida fresh waters have pH values between 7 and 8. Although the pH is the most common measured property or characteristic of a solution or a media, some growers and production managers still ignore to adjust the pH of their water when used for pesticide mixing. For better efficacy, anyone involved in pesticide mixing should use a pH meter. The pH affects the rate at which some herbicides are absorbed by plants. Adjusting the pH of the water allows the user to reduce the rates of herbicides without reducing their efficacy. The effectiveness of spray mixture in the spray tank can be affected by a number of variables. A significant impact on the

efficacy of many spray materials is the pH of the water used in the tank. In general, it is desirable to have the pH of the water below 7. Although several chemicals used today are effective at a wide range of pH conditions, many others can be subject to breakdown of the active ingredient at relatively high pH values. With extremely sensitive chemicals, this breakdown can begin between mixing and application. Sevin is among the common pesticides that lose their effectiveness quickly in alkaline (pH values greater than 7) solution. Therefore, it is recommended to reduce the pH of the water in the tank to increase the efficacy of some chemicals. Acidifying agents such as phosphoric acid and citric acid will lower the pH, but can drop it too low. Buffering agents, available from most distributors, will lower the pH to the desired range and help maintain it at that level. It is important to add the buffer to the spray tank water before pesticides are added. Glyphosate works better when ammonium sulfate is added to the spray tank at rates of 8.5 to 17 pounds for every 100 gallons of spray solution. Be careful when buffering tank mixes containing copper fungicides. Copper is more soluble in acidic water, and the resulting high concentrations will cause leaf and fruit burn. Aliette makes acid spray. Therefore, do not mix Aliette with copper. Always read the label of the buffering material as well as the label of the **pesticide**. It is also recommended to ask your chemical supplier for up-to-date information on the susceptibility of a material to hydrolysis. A good rule of thumb is to spray pesticide mixtures as soon after mixing as possible, mix only enough to treat the crop and do not allow the mixture to stand for a long period of time or overnight.

### **BROWN ROT**



Management of brown rot, caused by *Phytophthora nicotianae* or *P. palmivora*, is needed on both processing and fresh market fruit. While the disease can affect all citrus types, it is usually most severe on Hamlin and other early maturing sweet orange cultivars. Phytophthora brown rot is a localized problem usually associated with restricted air and/or water drainage. It commonly appears from mid-August through October following periods of extended high rainfall. It can be confused with fruit drop due to other causes at that time of the year. If caused by *P. nicotianae*, brown rot is limited to the lower third of the canopy because the fungus is splashed onto fruit from the soil. *P. palmivora* produces airborne sporangia and can affect fruit throughout the canopy.

Early season inoculum production and spread of *Phytophthora* spp. are minimized with key modifications in cultural practices. Skirting of the trees reduces the opportunity for soilborne inoculum to contact fruit in the canopy. The edge of the herbicide strip should be maintained just inside of the dripline of the tree to minimize the exposure of bare soil to direct impact by rain. This will limit rain splash of soil onto the lower canopy. Boom application of herbicides and other operations dislodge low-hanging fruit. Fruit on the ground becomes infected and produces inoculum of *P. palmivora* that can result in brown rot infection in the canopy as early as July while fruit are still green. The beginning stages of the epidemic are very difficult to detect before the fruit are colored and showing typical symptoms. Application of residual herbicides earlier in the summer may reduce the need for post-emergence materials later and minimize fruit drop throughout this early stage of inoculum production from fallen fruit.

Usually a single application of Aliette, Phostrol or ProPhyt before the first signs of brown rot appear in late July is sufficient to protect fruit through most of the normal infection period. No more than 20 lb/acre/year of Aliette should be applied for the control of all Phytophthora diseases. Aliette, Phostrol and ProPhyt are systemic fungicides that protect against postharvest infection and provide 60-90 days control. Copper fungicides are primarily protective but are capable of killing sporangia on the fruit surface and thus reducing inoculum. They may be applied in August before or after brown rot appearance and provide protection for 45-60 days. If the rainy season is prolonged into the fall, a follow-up application of either systemic fungicides at one-half of the label rate, or copper in October may be warranted. With average quality copper products, usually 2-4 lb of metallic copper per acre are needed for control.

Precautions should be taken during harvesting not to include brown rot-affected fruit in the field containers as this could result in rejection at the processing or packing facility.



### **Recommended Chemical Controls for Brown Rot of Fruit**

Pesticide	FRAC MOA <sup>2</sup>	Mature Trees Rate/Acre <sup>1</sup>	
Aliette WDG	P07	5 lb	
Phostrol	P07	4.5 pints	
ProPhyt	P07	4 pints	
copper fungicide	M01	Use label rate.	

<sup>&</sup>lt;sup>1</sup>Lower rates may be used on smaller trees. Do not use less than minimum label rate. <sup>2</sup>Mode of action class for citrus pesticides from the Fungicide Resistance Action Committee (FRAC) 2018. Refer to ENY624, Pesticide Resistance and Resistance Management, in the 2018-2019 Florida Citrus Production Guide for more details.

Rates for pesticides are given as the maximum amount required to treat mature citrus trees unless otherwise noted. To treat smaller trees with commercial application equipment including handguns, mix the per acre rate for mature trees in 250 gallons of water. Calibrate and arrange nozzles to deliver thorough distribution and treat as many acres as this volume of spray allows.

<u>For more information</u>, go to Florida Citrus Production Guide: Brown Rot of Fruit at: <a href="https://edis.ifas.ufl.edu/pdf/CG/CG022/CG022-Dkge3emz8a.pdf">https://edis.ifas.ufl.edu/pdf/CG/CG022/CG022-Dkge3emz8a.pdf</a>

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