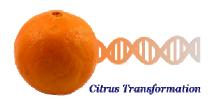
Transgenic Solutions at CREC

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Antimicrobial peptides and HLB/Canker/CVC



- No natural resistance to HLB in any commercially cultivated sweet orange, grapefruit or tangerine.
- Antimicrobial peptides have been shown to provide resistance to bacterial diseases.
 - LIMA gene to control Xylella
 fastidiosa, the causal organism for
 Pierce's disease in grapes. (Dr. Dennis Gray,
 MREC, UF/IFAS)
- Same gene can theoretically be used to combat all three diseases.

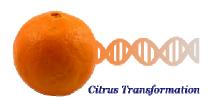






What are antimicrobial peptides?

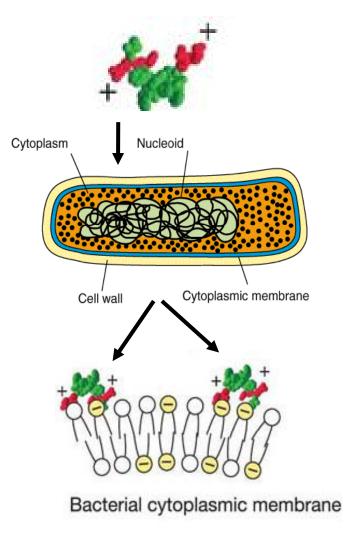
- Antimicrobial peptides are usually small proteins, usually 12 and 50 amino acids long.
- They form the first line of host defense against pathogenic infections and are a key component of the innate immune system
- Antimicrobial peptides are involved in the antimicrobial defense system among all classes of life.
 - Plants
 - Insects
 - Amphibians and
 - Mammals including humans





Mode of action of Antimicrobial peptides

- The net charge of antimicrobial peptides is positive. Also, they are hydrophobic and they are membrane active.
- The outer surface of bacteria is negatively charged.
- These peptides are mobilized shortly after microbial infection, and act rapidly to neutralize a broad range of microbes.
- The positively charged antimicrobial peptides bind to the negatively charged bacterial membrane
- The membrane is disrupted and antimicrobial peptides inflict damage that is difficult to repair.







Incorporation of antimicrobial peptides for disease resistance in citrus

Objectives

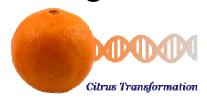
- Design codon optimized antimicrobial peptides genes for citrus to combat HLB and Canker.
- Target trans-protein in phloem tissue where HLB resides.
- Produce a large number of transgenic lines using *Agrobacterium* and protoplast mediated transformation.
- Challenge plants with disease causing bacteria to evaluate resistance.





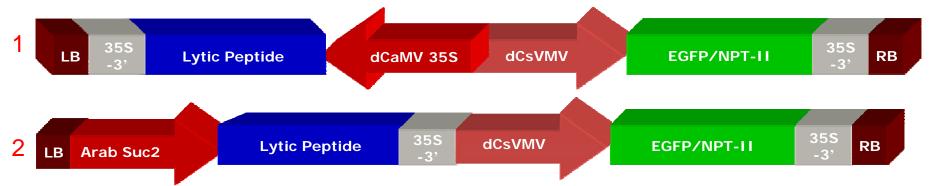
Antimicrobial gene(s) currently under evaluation

- AttacinE Lytic peptide gene from Hyalophora cecropia.
- CEAD Codon optimized cecropin A-cecropin D lytic peptide gene.
- CEMA Codon optimized cecropin A-melittin lytic peptide gene.
- CEME Codon optimized cecropin A-melittin lytic peptide gene (differs at the C terminus from CEMA).
- LIMA Lytic peptide gene obtained from Dr. Dennis Gray, MREC, UF/IFAS.
- PTA Codon optimized N terminally modified Temporin A gene.





Gene Construct(s) used for Transformation



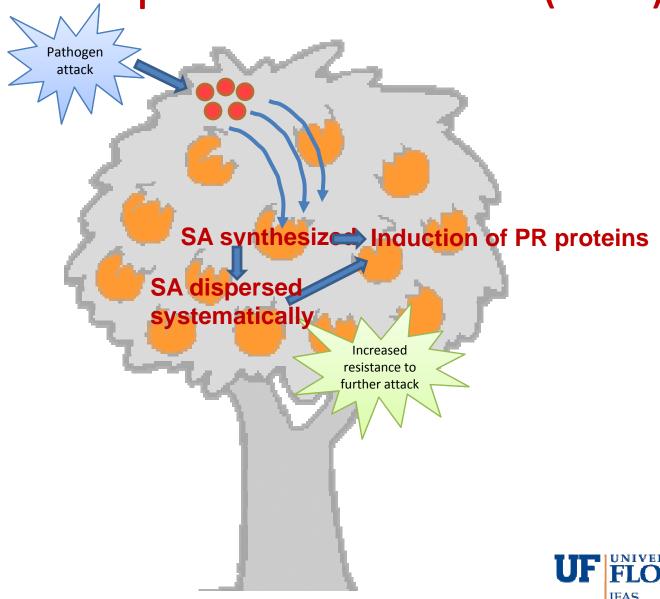
The antimicrobial gene(s) were driven by

- 1. Constitutive doubly enhanced 35s promoter or
- 2. Phloem specific sucrose synthase promoter.
- A green fluorescent protein/neomycin phosphotransferase II (EGFP/NPT II) bi-functional fusion gene under control of a cassava mosaic virus promoter was used to monitor and select transformed cells.
- A pBIN19 backbone was used to clone all gene constructs.
- Plasmid DNA was incorporated into Agrobacterium EHA105.





Systemic acquired resistance (SAR)







Genes for SAR

- SABP2 (Salicylic Acid-Binding Protein 2 gene from tobacco)
 - Isolated from the Tobacco plant.
 - High Affinity for SA.
 - It may be required to convert Methyl Salicylate to SA as part of the signal transduction pathways that activate systemic acquired resistance.





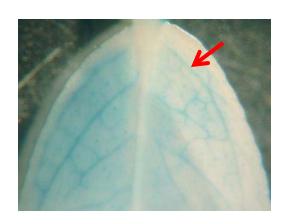
Genes for SAR

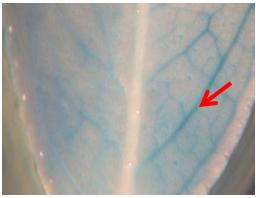
- NPR1 (Nonexpresser of PR Genes1 gene from Arabidopsis)
 - Isolated from the Model plant Arabidopsis thaliana.
 - NPR1 is a key regulator in the signal transduction pathway that leads to SAR.
 - Mediates the salicylic acid induced expression of pathogenesis-related (PR) genes and systemic acquired resistance.





Use of phloem specific promoters to restrict trans-protein in phloem tissues





GUS expression in citrus leaf phloem tissue using the Rice Sucrose Synthase promoter

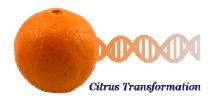
- HLB resides in the phloem.
- Targeting the trans-protein in the phloem resolves issues of the presence of the protein in the fruit and juice.
- Two phloem specific promoters are currently under evaluation
 - 1) Arabidopsis Sucrose synthase promoter and
 - 2) Rice Sucrose synthase promoter.





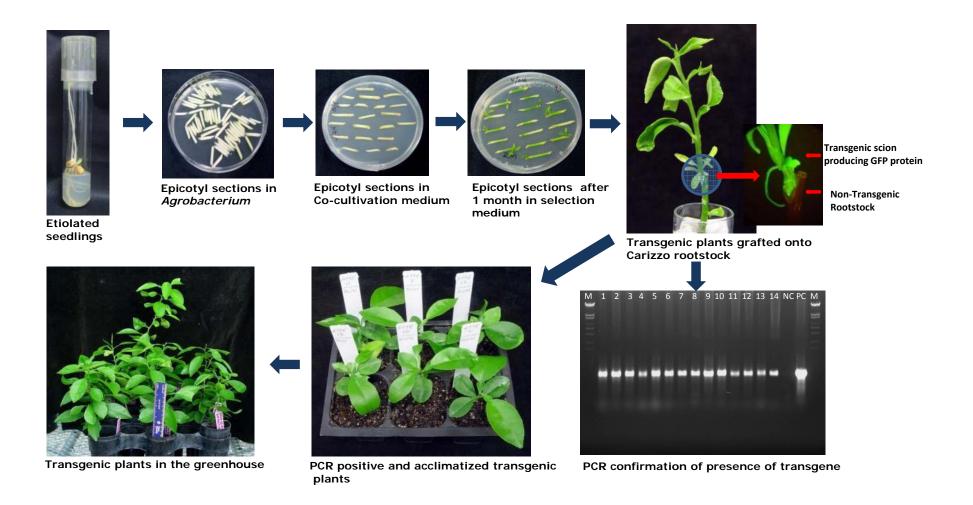
Genetic transformation of citrus

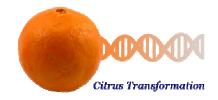
- Two methods now available
 - Agrobacterium mediated transformation
 - PEG mediated Protoplast transformation





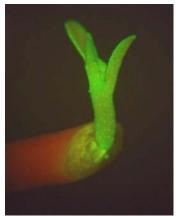
Agrobacterium Mediated Transformation



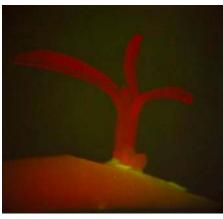




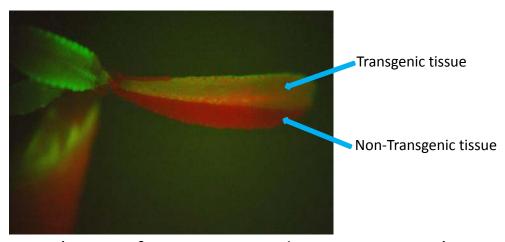
Selection of transgenic tissue



Transgenic tissue glows green!



Non-Transgenic tissue stays red



Chimeric tissue (a mix of transgenic and non transgenic)



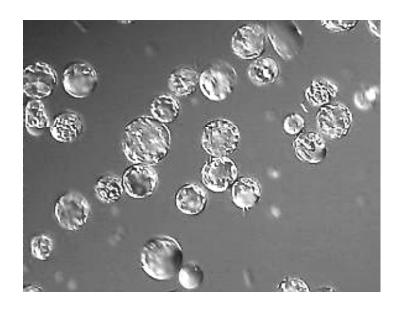


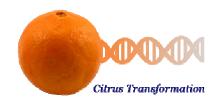
Citrus Protoplast Transformation

 A plant cell that had its cell wall completely removed using enzymatic means.



- Naked Plasmid DNA is incorporated into Protoplast DNA by Electroporation or PEG mediated transformation.
- Complete plants are regenerated from transformed protoplasts using Tissue culture techniques.





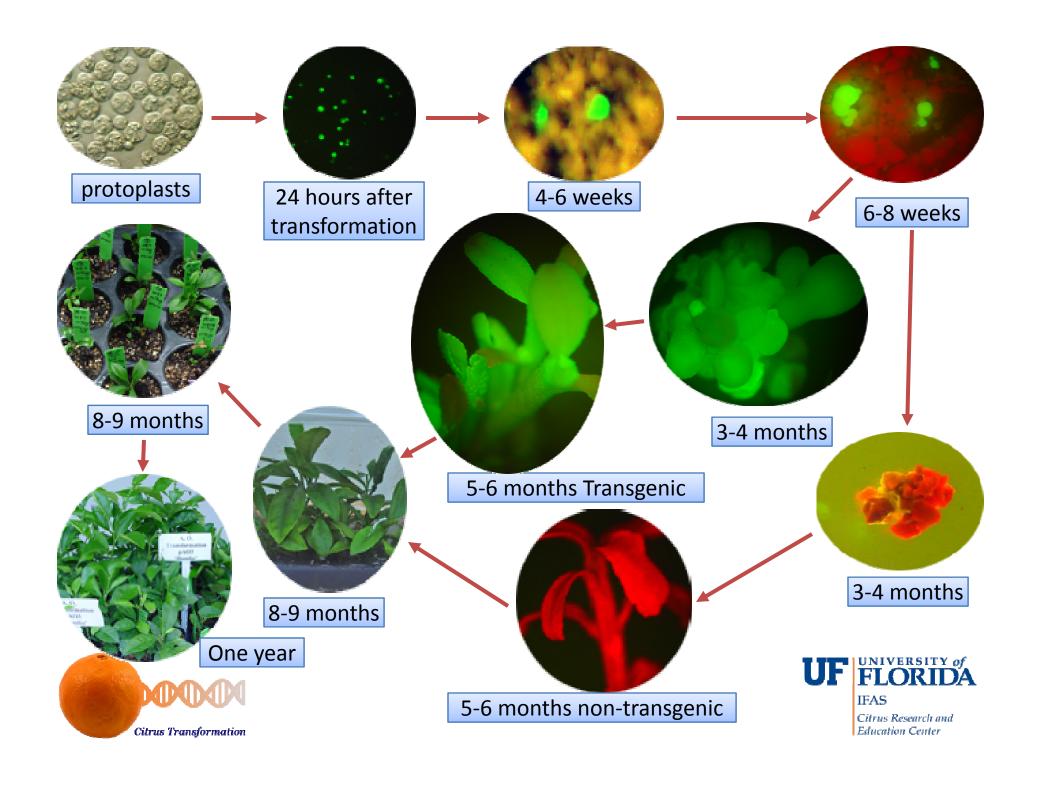


Protoplast transformation

- Can be used to transform polyembryonic seedless cultivars that are difficult to transform using Agrobacterium.
- Can be used to transform some polyembryonic mandarin cultivars that are often more recalcitrant to Agro-infection (i.e. W. murcott)
- Can be used to engineer plants not containing an antibiotic resistance gene.







Transgenic plant regeneration

Cultivar	Gene	No. of plants in soil	
Duncan	AttacinE	27	
Hamlin	AttacinE	15	
Misc Grapefruit	LIMA	45	
Valencia, Hamlin,OLL-8	LIMA	56	
Carrizo	LIMA	8	
Flame	LIMA-SN	10	
Misc Grapefruit	PTA	12	
Valencia, Hamlin, OLL8	CEMA	21	
Carrizo	CEMA	20	
Key Lime	CEMA	6	
Misc Grapefruit	CEME	18	
Hamlin	CEME	6	
Valencia	CEAD	14	
Carrizo	CEAD	12	
Carrizo	LIMA under AtSuc2 promoter	25	
Valencia, Hamlin, OLL-8	LIMA under AtSuc2 promoter	23	
Key Lime	LIMA under AtSuc2 promoter	17	
Misc Grapefruit	LIMA under AtSuc2 promoter	12	

Results with SAR genes

Cultivar	Gene construct	No of Transgenic	
		plants in soil	
Valencia, Hamlin,	35s - SABP2	30	
Flame			
Valencia, Hamlin	35s - NPR1	20	
Hamlin	AtSUC2 – SABP2	26	
Hamlin	AtSUC2 – NPR1	10	
Hamlin	35s – NPR1 + 35s - LIMA	2	





Rapid propagation of transgenics











Young budded transgenic plants for field testing







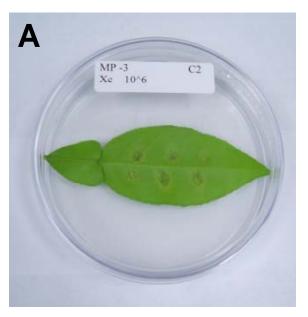
Transgenic trees ready for testing

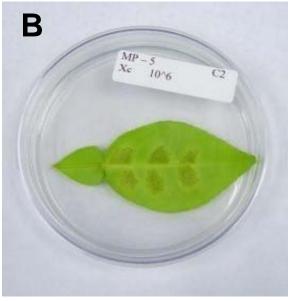


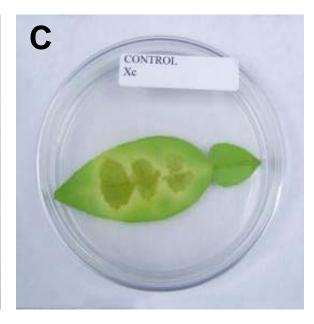




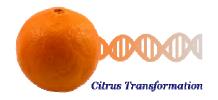
Canker (Xcc) assay on transgenic Duncan leaves containing the lytic peptide gene(s)







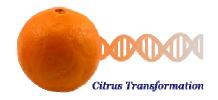
- A Mild tissue hypertrophy in transgenic LIMA leaf
- B Moderate tissue hypertrophy in transgenic Attacin leaf
- C Severe water-soaking and tissue hypertrophy in non-transgenic control leaf





qRT-PCR analysis of selected transgenic lines infiltrated with Xcc by the detached leaf assay method

Transgenic	AttacinE		LIMA	
line	Mean cT	Bacterial cells / mg tissue*	Mean cT	Bacterial cells / mg tissue
MP1	14.775	194975	18.285	18623
MP2	16.565	61453	18.362	17619
MP3	14.180	318000	26.625	70
MP4	15.935	90065	23.365	629
MP5	18.670	14369	14.040	324000
Control	15.560	115031	15.560	115031





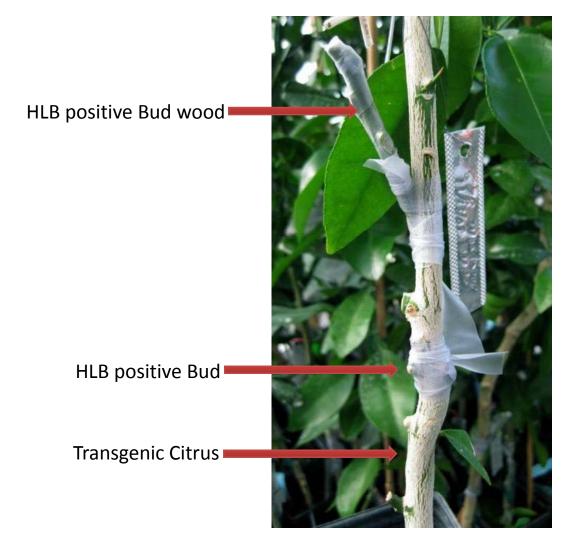
Transgenic plant challenge with HLB

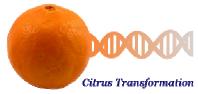
- Carried out in an approved secure facility.
- Transgenic plants graft challenged with HLB infected sweet orange budwood.
- Several Transgenic lines are currently under evaluation.
- Transgenic plants evaluated for HLB symptoms and infection verified by qRT-PCR.





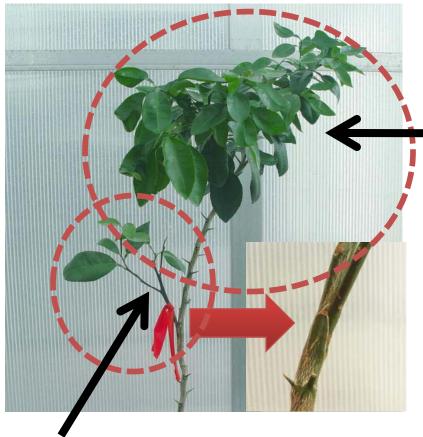
Graft and bud inoculation with HLB







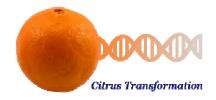
Transgenic plant challenge via grafting with HLB infected budwood



HLB infected budstick

Transgenic citrus plant after 13 months of inoculation with HLB + sweet orange bud stick

Explant	qRT-PCR using HLB specific primers
Transgenic plant	Negative
Budstick	Positive





HLB inoculation results

- Wide range of symptoms observed after inoculation with HLB + budwood
- General mottle on top of leaf
- Mottle and subsequent yellowing







THANKS!

- FCPRAC block grant entitled "SURVIVING HLB AND CANKER: GENETIC STRATEGIES FOR IMPROVED SCION AND ROOTSTOCK VARIETIES" - Fred Gmitter, Jude Grosser, Bill Castle and Gloria Moore
- USDA-CSREES
- UF/CREC Core Citrus Transformation Facility
- CREC Faculty and Staff





Psyllid control

- Production of Transgenic citrus plants to combat psyllids
- Duncan Grapefruit used as a model.
- Constitutive expression of a Snowdrop Lectin gene from the Snowdrop Plant (Galanthus nivalis)
- Several plants are currently being propagated for resistance studies.





Future approaches for genetic resistance studies

- Screen a large number of transgenic trees to select individuals with the highest level of resistance.
- Challenge putative resistant plants with psyllids.
- Field testing in a high disease pressure environment.







Cellulase Macerozyme



Citrus Transformation

Protoplast ring

